

### **Plant Science Research**

ISSN 0972-8546



# Bioprospecting of fungal endophytes from plants of the family Zingiberaceae and their pharmaceutical applications

Soumya Priyadarsini<sup>1</sup>, Deepa Biswas<sup>1</sup> and H.N. Thatoi<sup>2Ψ</sup> <sup>1</sup>Department of Botany, Kalinga University, Naya Raipur, Chhattisgarh 492101, INDIA <sup>2</sup>Department of Biotechnology, MaharajaSriram Chandra Bhanja Deo University, Baripada-757003, INDIA

#### ARTICLE INFO

Article history: Received : 11 October, 2022 Revised : 1 December, 2022 Accepted : 8 December, 2022

Keywords:

Endophytic Fungi, Zingiber plants, bioactive compounds, pharmaceutical applications.

#### ABSTRACT

Endophytic fungi are thought of as the hidden members of the microbial world that are widely recognised as a rich resource of potential chemical compounds with biological activities. The primary role of the bioactive substances produced by endophytes is to aid the host plants in withstanding abiotic and biotic stresses, which is ultimately beneficial to the survival of the host. Most of these endophytic fungi are from the Ascomycota, Oomycota, Zygomycota, and Basidiomycota groups. Research into endophytes has revealed that they have the potential to yield useful pharmaceuticals. Endophytic fungi are regarded as an under-utilized sourcethat hasn't been used enough to find new drugs and compounds. Endophytic fungi exhibit antibacterial, anticancer, immunomodulatory, antiviral, anti-diabetic, antioxidant, anti-inflammatory and other biological activities, which are being exploited for pharmaceutical potential of endophytic fungi from an important group of medicinalplant belonging to the family Zingiberaceace.

#### 1. Introduction

Endophytes are described as polyphyletic groups of microorganisms which may be actinomycetes, bacteria or fungi (Kusari et al., 2012). Endophytic fungi (EF) are a group of fascinating host-associated fungal communities that thrive asymptomatically the internal tissues of healthy plants without causing any diseases to the host (Amirita et al., 2012). They colonise the host either as obligate associates or as facultative associates with lower and higher plants (Deshmukh et al., 2006). Endophytes communicate information to higher plants and have evolved biochemical pathways that produce novel bioactive substances. Therefore, they offer chances for discovering products and processes with medical and biotechnological applications (Bielecka et al., 2022). In recent years, accumulated research on endophytic fungi has revealed their wide-ranging ecological distribution, biodiversity and multidimensional interactions with host plants and other microbiomes in the symbiotic continuum (Alam et al., 2021).

Corresponding author; Email: hn\_thatoi@rediffmail.com

© 2022 Orissa Botanical Society

It is believed that there are about one million different species of endophytic fungi that can be found in nature (Faeth and Fagan, 2002). According to Schultz, the endophytic fungi are mainly classified into three main ecological groups: (a) pasture or balansicaeousendophytic fungi; and (b) non pasture endophytic fungi; (c) mycorrhizal endophytic fungi (Faeth and Fagan, 2002).

Their affiliation can be compulsory or discretionary and does not, thus, cause direct damage to its host. Endophytes have dynamic relationship with its host plants, which include mutual relationship and antagonistic relationship. Plants primarily restrict endophyte growth and the endophytes undergo various mechanisms to steadily adapt into the living environment (Dudeja *et al.*, 2012). In order to maintain a sustainable symbiotic association, a range of compounds are produced by the endophytes such as gibberellins, indoleacetic acid, alkaloids, isocoumarin derivatives, quinones, chlorinated metabolites, flavonoids, terpenoids, phenol and phenolic acids which facilitates the

growth of the host plant and help them better survive in their environment. The presence of endophytes encourages plant nutrient absorption and thus contribute towards host plant's enhanced growth. Endophytes are considered as plant growth promoters by producing various broad range of phytohormones like auxins, gibberellins, IAA, cytokines etc. Besides plant growth, endophytes are known to induce growth in terms of increase in plant height, shoot diameter and shoot weight (Yates et al., 1997). Apart from plant growth factors, endophytes generate pharmaceutical agents which have biotechnological potential, like anti-tumor drugs (taxol), anti-fungal drugs (quercine), and various industrially important enzymes (Ting et al., 2008; Cao et al., 2004). Different enzymes are produced by endophytes include protease, amylase, pectinase, L-asparginase, chitinase, laccase, etc. Endophytes living inside medicinal or crop plants have confirmed the presence of most enzymes (Ayob et al., 2016).

During the past two decades, the endophytes are being explored for novel bioactive compounds. Various compounds generated via biosynthetic pathways belong to broad classes of compounds like phenols, tetralones, enniatins, steroids, isocoumarins. Endophytic organisms are actually a reservoir for the discovery of new compounds for development in medicinal and agricultural industries by producing various antibiotics, antioxidants, anti-parasitic and anti-cancer drugs. At present, main focus of exploring endophytes are the potential of these microorganisms in developing secondary metabolites. Since endophytic microbes may synthesis or even mimic the same substance produced by their host plants, bioprospecting of these microorganisms opens up avenues of discovering novel molecules (Venieraki *et al.*, 2017).

Notable species like ginger and turmeric belonging to family Zingiberaceae has long been recognized as a potential cure for a variety of conditions, including cramps, digestive issues, arthritis, fever, cough, and cold (Ma, 2012). Zingiberaceae plants have been extensively researched in the scientific literature because of their antimicrobial, antiapoptosis, anti-tumor, and anti-inflammatory and other biological activities, which can be attributed to the presence of several phyto-compounds in them (Karuppiah, 2012). Anti-microbial resistance (AMR) is a global health challenge on medicinal view point and there is a need for discovery of new antimicrobial compounds from natural sources. In this context, exploitation of fungal endophytes to synthesize novel antibiotics, especially the modified or somewhat similar with Zingiberaceae compounds has great prospect in medical application. The initial step for study about the biopotential of endophytic fungi is the isolation of some antagonistic fungal strains from non-symptomaticand healthy plant parts. Several researchers have successfully isolated and reported many fungal endophytes and evaluated their antibacterial potential. Antagonistic endophytic Penicillium sp. has been isolated from Curcuma longa, which exhibits inhibitory activity against human pathogenic bacteria such as Klebsiella pneumoniae and Pseudomonas aeruginosa (Rathod, 2013). The results of antagonistic activity of fungal strains towards examined pathogens were unique, demonstrating that fungal strains have varying capabilities in generating antimicrobial chemicals. Chemical compounds made by growing Pestalotiopsis vaccinii were thought to be new natural products made only by the strain itself (Wang, 2014; Wang, 2017). Research on application of fungal endophytes from the Zingiberaceae family is still in its infancy. However, earlier studies have revealed endophytic fungi (EFs) as new sources of bioactive chemicals with anticarcinogenic substances (Uzma et al., 2018). Therefore, the study of distribution, diversity, and biochemical properties of endophytes is a very important, which will help us to understand and improve plant fitness (Yuan et al., 2019). The present review provides a thorough explanation of the biological features of EFs from plants belonging to Zingiberaceae family, including their diversity, distribution, function along with pharmaceutical applications. Besides, the review also highlights the promising therapeutic uses and exhaustive pharmacological information of the species of Zingiberaceae.

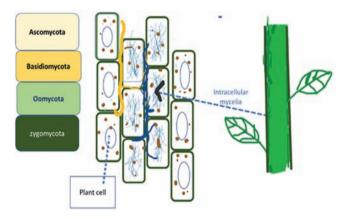


Fig. 1: Classification of endophytic fungi and their occurrence with in plant cell

### 2. Species and phytochemical diversity of family Zingiberaceae

The monocotyledonous family Zingiberaceae are comprised of a large number of medicinal, aromatic and ornamental plants, which are widely distributed throughout

the tropical and sub-tropical regions of the world (Ghosh et al., 2013) with greater species concentration in Southeast Asia (Jantan et al., 2003). The family is comprised of over 1200 species under 53 genera (Kress et al., 2002). As many as 178 species under 22 genera are reported from North Eastern and peninsular region of India (Jain and Prakash, 1995). The members of Zingiberaceae are both perennial or annual rhizomatous herbs. These aromatic ethnomedicinally important plants are characterised by the presence of volatile oils and oleoresins. The important members under the family Zingiberaceae having medicinaluses belongs to the genus Amomum, Curcuma, Alpinia Elettaria, Hedychium, Kaempferia and Zingiber (Prabhu et al., 2010). In general, the rhizomes and fruits belong to this group of plant are aromatic and are used as tonic and stimulant. These multipurpose plant family are the rich sources of natural products and chemicals thatare already being utilized as spices, medicines, dyes, perfume and food.

India, traditionally known as 'the spice bowl of the world', is the largest producer, consumer and exporter of ginger. Ginger has rich cultivar and genetic diversity in India, and the cultivars are generally named after localities of occurrence or cultivation. More than 50 cultivars possessing varying yield and quality parameters are grown in India. Kerala holds maximum diversity of ginger followed by Himachal Pradesh and Assam (Ravindran *et al.*, 2005). Disease-resistant and high-yielding ginger hybrids are the results of crop improvement programmes and Athira, Rajatha, Mahima, Suprabha, Suruchi, Suravi, Himagiri, Karthika, Varada are some popular varieties developed in India.

Endophytic fungi live in rhizomes of plants of ginger family and the metabolites that these fungi make may have an effect on the bioactivity of ginger extracts and the quality of the spice. These metabolites are not removed by washing, so they may also have an effect on human health. These metabolites are produced against host-specific phytopathogens and may demonstrate antibacterial, antifungal, cytotoxic, anticancer, antioxidant, and antiinflammatory activities.

Though fungi are capable of colonising intercellular or intracellular regions of host plant, they are more likely to do so at the plant's roots than at its leaves or stems. Apoplastic fluid from the host is the primary nutrition supply for colonisation in aerial organs, allowing for healthy reproduction of EFs (Schulz *et al.*, 2002). During an infection, fungi form three distinct types of interactions with their hosts: communalistic (non-beneficial/virulent endophytes), mutualistic (beneficial endophytes) and pathogenic (virulent pathogens). The nature of these interactions differs depending on the physiological status of the host plants or the specific conditions that they are exposed to. *Alpinia, Amomum, Curcuma, Elettaria, Hedychium, Kaempferia* and *Zingiber* are the interesting genera of Zingiberaceae which contain aromatic spice plants with very high medicinal values (Joy *et al.*, 1998). Among them, few species are cultivated but majority of the species occur in the wild. Each of the above-mentioned genera are briefly discussed below.

#### 2.1. Genus Alpinia

Alpinia Roxb. is the most widespread and largest genus belonging to subfamily Alpinioideae under the Zingiberaceae with about 230 species. Members of this genus are mainly distributed in India, Indonesia, Arabic gulf areas, Egypt, Malaysia, Japan, southeast Asia, the Pacific, Samoa, the Caroline Islands, Australia and northern New South Wales, China and Sri Lanka (Larsen et al., 1998; Smith, 1990). Most of the species grow in open and sunny places, low- to mid-elevation forests and brushwood. In India, the species of these genus are found in the Himalaya and Southern region of Western Ghats (Khare et al., 2007). These are perennial herbs with oblong-lanceolate leaves, tuberous root, greenish white flowers. (Gupta et al., 2010). While the rhizomes are source of essential oils (Jirawan et al., 2005), the young shoots and flowers are used as a spice (Arambewela et al., 2006) and condiments. In Asian countries, Alpinia species are used in cooking (A. galanga) and medicinal purposes (e.g., A. officinarum Hance). The plant is mostly used in the traditional system of medicine viz. Ayurveda, Unani, Chinese and Thai folk medicines (Yang et al., 1999). The active phytochemical constituents of these plants support medicinal and pharmacological properties.

#### 2.2. Genus Curcuma

*Curcuma* is a genus with some 120 species occurring from Indochina, Malaysia, India, northern Australia and Thailand (Leong, 2007; Larsen, 1996; Maknoi, 2006). It is a genus of perennialherbs whose rhizomes are fleshy, aromatic and usually varies from light brown to different shades of yellow (Maknoi, 2007). India is the top producer of *Curcuma* rhizomes followed by Southeast Asia, Thailand, Central and Latin America, and Taiwan.

Many of its members have medicinal uses such as treating digestive disorders including dyspepsia and colic due to its anti-inflammatory, anti-viral, anti-bacterial, anti-fungal, and anti-oxidant properties. The anti-cancerous activity from extract of *Curcuma longa* is under trials. *Curcuma* is of great economic importance, with *Curcuma longa* as the top notched in the genus along with *C. amada*, *C. angustifoli*a and *C. zedoria*, *C. caesia* are common species.

#### 2.3. Genus Zingiber

The genus Zingiber is comprised of about 85 species of herbs mostly grown in Asia, Africa, central and south America (Sabulal et al., 2006). These plantae prefer moist, tropical conditions (Javashree et al., 2015). Phytochemical investigation of the rhizomes of several Zingiber species has revealed that they possess various pharmacological and physiological effects due to the presence of bioactive compounds such as gingerols, shogaols, diarylheptanoids, phenylbutenoids, flavanoids, diterpenoids and sesquiterpenoids (Sivasothy et al., 2011). In addition, shogaols, dehydrated gingerol derivatives, are the predominant pungent constituents in dried ginger (Jiang et al., 2006). These plants are used as common ingredients in traditional medicines. The rhizomes have been used successfully to treat a variety of medical conditions, including gastrointestinal distress, motion sickness, vomiting, epilepsy, bronchitis, common cold, bruises, wounds, liver complaints, rheumatism, muscular pains, atherosclerosis, migraine headaches, high cholesterol, ulcers, and stomach discomfort (Shukla et al., 2007). Phenolic compounds in ginger root, especially gingerols, have been shown to have chemopreventive effects linked to their antioxidant and anti-inflammatory properties (Shukla et al., 2007).

#### 2.4. Genus Hedychium

The genus *Hedychium* has 87 species, which have worldwide distribution (Plant list, 2010). *Hedychium spicatum*, found from Himachal Pradesh to Arunachal Pradesh, is useful in the treatment of liver complaints, fevers, vomiting, diarrhoea, inflammation, pains and snake bite. The rhizomes of *H. spicatum* are reported to harbour twenty-eight fungal strains and about 84% of endophytic fungi isolated belonged to the phylum Ascomycota. Endophytic fungal genera such as *Fusarium* and *Penicillium* were found to be common to all the plant parts. *Hansfordia biophila* is an endophytic fungus isolated from *Hedychium acuminatum* that produces tannin (Hastuti *et al.*, 2018).

#### 2.5. Genus Elettaria

The genus *Elettaria* is represented by few species of perennial herbs with rhizomes. They grow in India, Sri Lanka, Malaysia, and Indonesia (Wills, 1967). In India, the species are indigenous to the most southern tropical evergreen forests of the Western Ghats of India (Ravindran, 2002). Among them, Cardamom (*Elettaria cardamomum*), also known as Indian cardamom or small cardamom, has been used worldwide for culinary purposes and the older forms of medicine. Essential oils and other high-value antioxidant and gastroprotective bioactive metabolites from its fruits are primarily responsible for its unique aroma and function as a functional food as well as a nutraceutical and medicine (Hamzaa and Osman, 2012).

#### 2.6. Genus Amomum

Amomum, another genus of Zingiberaceae, is represented by over 150 species, mostly in Asia and Oceania (Cai et al., 2021). They are perennial herbs with distichous leaves and elongating pseudo-stems (Syazana, 2018). Volatile oils, as one of the essential components of the genus Amomum, have been extensively studied. Till date, more than 160 non-volatile compounds have been isolated from this genus, including flavonoids, terpenoids, diarylheptanoids, etc. In China, India, Thailand and Nepal, 16 species of the genus Amomum are utilised in traditional medicine. Recent exploratory research on the pharmacological qualities of the Amomum species revealed their efficacy as anti-inflammatory agents (Jin et al., 2016), cure for stomach diseases (Kumar et al., 2014), cancer (Zhang et al., 2015), hepatopathy diseases (Kim et al., 2015) and malaria (Heilmann et al., 2001).

#### 3. How endophytic fungi colonise within host plants

It is well established that endophytic fungus colonisation is not a random event and happens because of chemotaxis, or specialised molecules produced by the host plant. At the same time, different types of secondary metabolites like saponin and essential oils from medicinal plants, are made as a way to fight off pathogens which acted as barriers against the colonisation of endophytic fungi. To circumvent this, the endophytic fungi must secrete the corresponding detoxifying enzymes such as cellulases, lactases, xylanases and proteases for the degradation of these secondary compounds before they reach the defence systems of the host-plants. Once within the tissues of a host-plant, endophytic fungi assume a quiescent (latent) state, either for the entire lifetime of the host plant (neutralism) or for an extended period of time (mutualism or antagonism), until environmental conditions are favourable for endophytic fungi or the ontogenetic state of the host changes to the fungi's advantage (Sieber, 2007).

Different interactions have been developed between endophytic fungi and their host plants through a distinct fungus-host interaction during the lengthy period of coexistence and evolutionary processes: (i) a continuum of mutualism, (ii) antagonism, and (iii) neutralism. The genetic background, nutrient level, and ecological habitats of medicinal host plants are thought to be pressure-choice factors on the population structure of endophytic fungi, which in turn confer some kinds of benefits, such as induced growth, increased resistance to disease, and/or herbivores, as well as accumulated bioactive components (Firakova *et al.*, 2007). For this reason, the mutual relationship between endophytic fungi and their host plants might impose specific effects on the formation of particular types of bioactive substances which have been explored for human benefits. To the best of our knowledge, there has been very little research on the commercialization of fungal endophytes isolated from Zingiberaceous plants.

### 4. Diversity of endophytes associated with the plants of the family Zingiberaceae

Endophytic fungi associated with different species of Zingiberaceae have been isolated from meristems, leaves, roots, stems, rhizomes by different workers. Generally, they are isolated by surface sterilization followed by culturing them from either crushed tissue extract or culturing through plant tissues on suitable media (Hata *et al.*, 2008). Members of family Zingiberaceae have been explored for a diverse group of endophytic fungi. Eleven fungal strains have isolated from *Curcuma longa* plant by Septiana (2017). About 33 fungal isolates have been isolated from six species of Zingiberaceae viz. *Zingiber officinale, Curcuma domestica, Kaempferia rotunda, Curcuma xanthorrhiza, Curcuma mangga* and *Curcuma zedoaria* (Praptiwi *et al.*, 2016).

Members of 10 fungal genera have been reported from Alpinia officinarum by Shubin et al. (2014). Endophytic fungi such as Trichoderma sp., Mycelia sterilia, Penicillium sp., Alternaria sp., Penicillium sp., Fusarium sp., Aspergillus sp., Bipolaris sp. and Nigrospora sp. have been isolated from Hedychium coronarium by Uzma et al. (2016). Similarly, species of Colletotrichum, Fusarium, Bipolaris, Pithomyces, Mucor, Alternaria, Mycelia, Rhizopus, Cladosporium are reported from Hedychium flavescens (Uzma et al., 2016). Bussaban et al. (2001) reported occurrence of Eupenicillium crustaceum, Fusarium spp., Glomerella spp., Phomopsis spp., Phyllosticta spp., Pyricularia spp., Talaromyces flavus, taxa of Xylariaceae, Mycelia sterilia from Amomum siamense.

### 5. Isolation of endophytic fungi from species of Zingiberaceae

Surface sterilization is the most needed technique which is used for the isolation of endophytic fungi from ginger family. Surface sterilized by immersing in 70% ethanol for 1 min. Then rinse the samples three times with sterile distilled water and soak them again in hypochlorite solution (NaOCl) 0.5% for 5 min and again rinse them in sterile water for six times. Allow the samples to become surface-dry by putting them on sterilized filter paper for 12 hours and it should be carried out in biosafety cabinet (Hallmann *et al.*, 2006). Place all the sterilized segments of each plant part on PDA containing chloramphenicol antibiotic (0.5 g/l) or streptomycin to inhibit the bacterial growth. Seal the Petri dishes with parafilm to prevent desiccation of the medium and incubate them in dark at 28 °C for 7-21 days. Continuously monitor the fungal growth. When the fungi grow out from the plant organ, then cut the hyphal tip arising from the fungal colony and transfer them to fresh PDA to enhance the normal sporulation for better identification. Every precaution should be practiced while performing the endophytic fungal isolation, as there are many chances of existence of epiphytic fungi during isolation.

Fungal identification was accomplished by observing their morphological characteristics. Besides, PCR and DNA sequencing are novel techniques employed to identify endophytic fungus. Identification of fungal endophytes can best be accomplished by amplification of Internal Transcribed Spacer (ITS) regions, which are repeating units of DNA encoding ribosomal RNA.

## 6. Biopharmaceutical potential of endophytic fungi from Zingiberaceae

Endophytic fungi are regarded as rich sources of numerous bioactive secondary metabolites and phytohormones that promote plant growth and enable plants to withstand biotic or abiotic stresses (Tan and Zou, 2001; Chadha et al., 2014). Apart from the benefits to the host plant, the endophytes have also proved as an outstanding source of SMs and bioactive antimicrobial products. Endophytic fungi can biosynthesize some of the most important commercially exploited SMs, including antibiotics, anticarcinogenic, cytotoxins, insecticides, and allelopathic compounds (Schneider et al., 2008). They have a lot of commercial potential in the pharmaceutical, medical, agricultural, nutraceutical, cosmetic, flavour, and fragrance industries. This makes EFs an interesting topic for endophytism research (Hyde and Soytong, 2008). The potential of endophytic microorganisms as untapped resources for innovative drugs has aroused a great deal of attention (Strobel, 2002). The ginger species are well-studied for their biological properties such as anti-microbial, antioxidant, anti-tumor, antihyperglycemic, anti-hyperuricemic, anti-inflammatory, anti-larvae, anti-aging and functional properties as a topical medicine attributed to presence of alkaloids, flavonoids, glycosides, phenols, saponins, steroids, tannins, and terpenoids (Juwita et al., 2018).

Hypomontagnella monticulosa (Zg 15SU) isolated from Zingiber griffithii showed potent cytotoxic activity (Lutfia *et al.*, 2021). Bioactive compounds such as acyclocitral, cembrene A, laurenan-2-one, sclareol, 2Z, 6E farnesol, cembrene, isocomene and  $\alpha$ -curcumene obtained

from endophytic fungi Arthrinium sp. (MFLUCC16-1053) account for their antimicrobial and antioxidant activities, which were isolated from Zingiber cassumunar (Pansanit et al., 2018). Zingiber zerumbet rhizomes harbour Fusarium solani and F. oxysporum which has antipythium activity (Keerthi et al., 2016). Similarly, the phenolic compounds produced by the fungal endophyte Bipolaris specifera, Alternaria tenuissima, Aspergillus terreus, Nectria haematococca and Fusarium chlamydosporum from Zingiber nimmonii have potential antioxidant properties (Das et al., 2017). Eurotium sp. isolated from Curcuma longa produces asparginase that has anticancer as well as antimicrobial properties (Jalgaonwala et al., 2010; Jalgaonwala et al., 2014). The antimicrobial and oxidant activity of 33 fungal isolates obtained from Zingiber officinale, Curcuma domestica, Kaempferia rotunda, Curcuma xanthorrhiza, Curcuma mangga and Curcuma zedoaria have been reported (Praptiwi et al., 2016). MM2 compound isolated from endophytic fungi Aspergillus terreus (RTN3) from young stems of Alpinia chinesis has shown potent anticancer activity and inhibitory effects on Staphylococcus aureus and methicillin-resistant Staphylococcus aureus (Vo et al., 2020). The ability of endophytic fungus Trichoderma afroharzianum isolated from Hedychium coronarium in producing chitinase has been reported (Munir *et al.*, 2019). The major bioactive compounds synthesized from endophytic fungi and their bioactivities are summarized below in Table 1.

Although the pharmacological activities of Zingiberaceae species have been extensively described, not much is known about bioprospecting of their endophytic microorganisms. Endophytic mycoflora are regarded as an exceptional pharmaceutical bioresource as they produce a wide range of substances known as secondary metabolites (Strobel, 2003). As bioactive compounds produced by endophytic community are believed to be regulated by the positive effect of plants, these plants with medicinal characteristics are regarded as a significant resource for studying fungal endophytes (Venkateswarulu et al., 2018). Endophytes have been found to be good sources of biologically active products that are interesting for certain health care uses (Strobel et al., 2001; Wiyakrutta et al., 2004). Apart from the benefits to the host plant, the endophytes have also proved as an outstanding source of secondary metabolites and bioactive antimicrobial products. Although there are many different endophytic fungi of documented pharmaceutical benefits isolated from family Zingiberaceae, we have complied a list of some of the most important ones in Table 1.

#### Table 1.

Sl. No.	Host plant	Endophytic fungi	Chemical compound / Enzyme	Pharmaceutical activities / Industrial application	References
1	Zingiber griffithii	Hypomontagnella monticulosa Zg15SU	Sesterpenoid	Anticancer	Lutfia <i>et al.</i> , 2021
2	Zingiber nimmonii	Bipolarisspecifera, Alternaria tenuissima, Aspergillus terreus, Nectria haematococca and Fusarium chlamydosporum	Phenolic compounds	Antioxidative	Das <i>et al.</i> , 2017
3	Zingiber cassumunar	Arthrinium sp. MFLUCC16-1053	Bcyclocitral, 3Ecembrene A, laurenan- 2- one, sclareol, 2Z, 6Efarnesol, cembrene, βisocomene and γcurcumene	Antibacterial and antioxidant activities	Pansanit <i>et</i> <i>al.</i> , 2018

List of endophytic fungi isolated from plants of family Zingiberaceae showing their pharmaceutical and industrial activities.

Bioprospecting of endophytic fungi from Zingiberacea	Bioprospecting	of	endophytic	fungi	from	Zingiberacea
------------------------------------------------------	----------------	----	------------	-------	------	--------------

Sl. No.	Host plant	Endophytic fungi	Chemical compound / Enzyme	Pharmaceutical activities / Industrial application	References
4	Zingiber zerumbet	Fusarium solani and F. oxysporum		Antipythium activity	Keerthi <i>et al.</i> , 2016
5	Curcuma longa	Eurotium sp.	Asparginase	Anticancer	Jalgaonwala <i>et al.</i> , 2014
6	Curcuma longa	Fungal isolates	-	Antibaterial and antifungal	Jalgaonwala et al., 2010
7	Alpinia calcarata	Cylindrocephalum sp.	Amylase	-	Sunitha <i>et al.</i> , 2012
8	Elettaria sp.	<i>T. harzianum</i> and <i>T. atroviride</i>	Antifungal enzyme	Antifungal	Munir <i>et al.</i> , 2019
9	Alpinia chinesis	Aspergillus terreus	MM1	Anticancer	Vo et al., 2020
10	Alpinia chinesis	Aspergillus terreus	MM1	Antimicrobial against Staphylococcus aureus and Methicillin-resistant Staphylococcus aureus (MRSA)	Vo et al., 2020
11	Elettaria sp.	Trichoderma harzianum	-	Antagonistic activity	Munir <i>et al.</i> , 2019
12	Elettaria sp.	Trichoderma atroviride	IAA production	-	Munir <i>et al.</i> , 2021
13	Hedychium coronarium	Trichoderma afroharzianum	Chitinase	-	Munir <i>et al.</i> , 2019
14	Hedychium coronarium	Trichoderma afroharzianum	-	Fusarium oxysporum	Munir <i>et al.</i> , 2019
15	Curcuma longa	Arthrobotrys foliicola, Cochliobolus kusano, Daldinia eschscholz, Fusarium oxysporum, Fusarium proliferatum, Fusarium solani, Fusarium verticillioides, Phanerochaetechrys osporium, Phaeosphaeria ammo philae		Inhibit growth of thehistamineproducing bacteria	Septiana <i>et</i> <i>al.</i> , 2017
16	Zingiber officinale	Curvularia affinis	-	Antimicrobial against Fusarium oxysporum	Ginting <i>et al.</i> , 2013
17	Zingiber officinale, Curcuma domestica, Kaempferia rotunda, Curcuma xanthorrhiza, Curcuma mangga and Curcuma zedoaria	33fungal isolates	-	Anti-oxidant and antimicrobial	Praptiwi <i>et</i> <i>al.</i> , 2016
18	Hedychium coronarium	Endophytic fungal strain	Staphylococcus aureus	Antimicrobial	Lutfia <i>et al.</i> , 2018

#### 7. Conclusion and future prospective

The current study shows that endophytic fungal metabolites are important targets for finding and making new drugs. Currently, the quest for novel biologically active metabolites from plant endophytes has broadened in light of the changing perspectives on medicinal substances and the rising demand for non-hazardous medications. We are losing the battle against therapeutic antimicrobial drugs due to development of resistance over the period of time. Endophytes, on the other hand, are a viable alternative since they contain an abundance of unique bioactive chemicals with virtually limitless potential biological activities. Numerous studies have reported novel, beneficial bioactive compounds exhibiting other biological properties, such as anti-diabetic, anti-inflammatory, antiprotozoal, antituberculosis, insecticidal, immune-modulatory, antiviral, anticancer activities, anthelmintic, etc., that were successfully isolated from endophytic fungi. Since the turn of the century, endophytic fungi have been the subject of intense study in the pharmaceutical industry due to their abundance and pervasive distribution. Although bioactive chemicals and the usage of certain medicinal plants in traditional medicine have been extensively explored, little is known about the bioactivities of its associated endophytic fungus. Despite this, limited research has been conducted on the valuable bioactive compounds from endophytic fungi. Thus, the screening of endophytic mycoflora for the possible SMs synthesis can help in the establishment of their pharmaceutical functions for sustainable human health and effective against antibiotic resistance.

#### References

- Alam, B., Li, J., Ge, Q., Khan, M.A., Gong, J., Mehmood, S., Yuan, Y. and Gong, W. (2021). Endophytic Fungi: From Symbiosis to Secondary Metabolite Communications or Vice Versa? Front. Plant Sci. 3060.
- Amirita, A., Sindhu, P., Swetha, J., Vasanthi, N.S. and Kannan, K.P. (2012). Enumeration of endophytic fungi from medicinal plants and screening of extracellular enzymes. World J. Sci Technol. 2(2): 13-19.
- Anisha, C. and Radhakrishnan, E.K. (2017). Metabolite analysis of endophytic fungi from cultivars of *Zingiber* officinale Rosc. identifies myriad of bioactive compounds including tyrosol. 3 Biotech. 7(2): 1-10.
- Arambewela, L., Wijesinghe, A. and Warnasuriya, D. (2006). *Alpinia galanga*. Sri Lankan medicinal plant monographs and analysis. 10, Industrial Technology Institute and National Science Foundation, Colombo, Sri Lanka.
- Ayob, F.W. and Simarani, K. (2016). Endophytic filamentous fungi from a *Catharanthus roseus*: Identification and

its hydrolytic enzymes. Saudi Pharm J. 24(3): 273-278.

- Bielecka, M., Pencakowski, B. and Nicoletti, R. (2022). Using next-generation sequencing technology to explore genetic pathways in endophytic fungi in the syntheses of plant bioactive metabolites. Agriculture 12(2):187.
- Cai, R., Yue, X., Wang, Y., Yang, Y., Sun, D., Li, H. and Chen, L. (2021). Chemistry and bioactivity of plants from the genus *Amomum*. J Ethnopharmacol. 281:114563.
- Cao, L., Qiu, Z., Dai, X., Tan, H., Lin, Y. and Zhou, S. (2004). Isolation of endophytic actinomycetes from roots and leaves of banana (*Musa acuminata*) plants and their activities against *Fusarium oxysporum* f. *cubense*. World J. Microbiol. Biotechnol. 20(5): 501-504.
- Chadha, N., Mishra, M., Prasad, R. and Varma, A. (2014). Root endophytic fungi: research update. J. Biol. Life Sci. USA. 5(2):135-158.
- Das, M., Prakash, H. S. and Nalini, M. S. (2017). Antioxidative properties of phenolic compounds isolated from the fungal endophytes of *Zingiber nimmonii* (J. Graham) Dalzell. Front. Biol. 12(2): 151-162.
- Deshmukh, S., Huckelhoven, R., Schafer, P., Imani, J., Sharma, M., Weiss, M., Waller, F. and Kogel, K. H. (2006). The root endophytic fungus *Piriformospora indica* requires host cell death for proliferation during mutualistic symbiosis with barley. Proceedings of the Nat. Acad. Sci. 103(49): 18450-18457.
- Dudeja, S. S., Giri, R., Saini, R., Suneja Madan, P. and Kothe, E. (2012). Interaction of endophytic microbes with legumes. J. Basic Microbiol. 52(3): 248-260.
- Faeth, S. H. and Fagan, W. F. (2002). Fungal endophytes: common host plant symbionts but uncommon mutualists. Integr. Comp. Biol. 42(2): 360-368.
- Firakova, S., Sturdikova, M. and Muckova, M. (2007). Bioactive secondary metabolites produced by microorganisms associated with plants. Biologia 62(3): 251-257.
- Ginting, R. C. B., Sukarno, N., Widyastuti, U. T. U. T., Darusman, L.K. and Kanaya, S. (2013). Diversity of endophytic fungi from red ginger (*Zingiber officinale* Rosc.) plant and their inhibitory effect to *Fusarium* oxysporum plant pathogenic fungi. Hayati J. Biosci.20(3): 127-137.
- Gupta, R.K. (2010). Medicinal and aromatic plants. CBS Publishers and Distributors 234: 499.
- Hallmann, J., Berg, G. and Schulz, B. (2006). Isolation procedures for endophytic microorganisms. *In*: Microbial root endophytes. Springer, Berlin, Heidelberg., pp. 299-319.

- Hammerschmidt, L., Ola, A., Mueller, W. E., Lin, W., Mandi, A., Kurtan, T. *et al.*, (2015) Two new metabolites from the endophytic fungus *Xylaria* sp. isolated from the medicinal plant *Curcuma xanthorrhiza*. Tetrahedron Letters. 56(10): 1193-1197.
- Hamzaa, R. G. and Osman, N. N. (2012). Using of coffee and cardamom mixture to ameliorate oxidative stress induced in ã-irradiated rats. Biochem. Anal. Biochem. 1(113): 2161-1009.
- Hardoim, P. R., van Overbeek, L. S. and van Elsas, J. D. (2008). Properties of bacterial endophytes and their proposed role in plant growth. Trends Microbiol. 16(10): 463-471.
- Hastuti, U. S., Asna, P. M. A. and Rahmawati, D. (2018). Histologic observation, identification, and secondary metabolites analysis of endophytic fungi isolated from a medicinal plant, *Hedychium acuminatum* Roscoe. *In:* AIP Conference Proceedings. 2002(1), pp. 020070.
- Hata, K. and Sone, K. (2008). Isolation of endophytes from leaves of *Neolitsea sericea* in broadleaf and conifer stands. Mycoscience. 49(4): 229-232.
- Heilmann, J., Brun, R., Mayr, S., Rali, T. and Sticher, O. (2001). Minor cytotoxic and antibacterial compounds from the rhizomes of *Amomum aculeatum*. Phytochemistry. 57(8):1281-1285.
- Hyde, K. D. and Soytong, K.(2008). The fungal endophyte dilemma. Fungal Divers. 33(163): 173.
- Jalgaonwala, R. E. and Mahajan, R. T. (2014). Production of anticancer enzyme asparaginase from endophytic *Eurotium* sp. isolated from rhizomes of *Curcuma longa*. Eur. J. Exp. Biol., 4(3): 36-43.
- Jalgaonwala, R. E., Mohite, B. V. and Mahajan, R. T. (2010). Evaluation of endophytes for their antimicrobial activity from indigenous medicinal plants belonging to North Maharashtra region India. Int. J. Pharm. Biomed. Res. 1(5): 136-141.
- Jayashree, E., Kandiannan, K., Prasath, D., Pervez, R., Sasikumar, B., Senthil Kumar, C. M., Srinivasan, V., Suseela Bhai, R. and Thankamani, C.K. (2014). Ginger. ICAR-Indian Institute of Spices Research, Kozhikode.
- Jiang, H., Xie, Z., Koo, H. J., McLaughlin, S. P., Timmermann, B. N. and Gang, D. R. (2006). Metabolic profiling and phylogenetic analysis of medicinal *Zingiber* species: Tools for authentication of ginger (*Zingiber officinale* Rosc.). Phytochemistry. 67(15): 1673-1685.
- Jin, M., Choi, J.K., Choi, Y.A., Kim, Y.Y., Baek, M.C., Lee, B. H., Jang, Y. H., Lee, W. J., Lee, S. J., Lee, H. S. and

Park, E. K. (2016). 1, 2, 4, 5-Tetramethoxybenzene suppresses house dust mite-induced allergic inflammation in BALB/c mice. Int. Arch. Allergy Immunol. 170(1): 35-45.

- Joy, P. P., Thomas, J., Mathew, S. and Skaria, B. P. (1998). Zingiberaceous medicinal and aromatic plants. Aromatic and Medicinal Plants Research Station, Odakkali, Asamannoor.
- Juwita, T., Puspitasari, I. M. and Levita, J. (2018). Torch ginger (*Etlingera elatior*): A review on its botanical aspects, phytoconstituents and pharmacological activities. Pak. J. Biol. Sci. 21(4): 151-165.
- Keerthi, D., Aswati Nair, R. and Prasath, D. (2016). Molecular phylogenetics and anti-Pythium activity of endophytes from rhizomes of wild ginger congener, *Zingiber zerumbet* Smith. World J. Microbiol. Biotechnol. 32(3): 1-11.
- Khare, C. P. (2007). Alpinia galanga An important medicinal plant: A review. A Dictionary of Indian Medicinal Plant. Springer Science + Business Media, LLC, New York
- Kim, H. G., Han, J. M., Lee, J. S., Suk Lee, J. and Son, C. G. (2015). Ethyl acetate fraction of *Amomum xanthioides* improves bile duct ligation-induced liver fibrosis of rat model via modulation of profibrogenic cytokines. Sci. Rep. 5(1): 1-11.
- Kress, W. J., Liu, A. Z., Newman, M. and Li, Q. J. (2005). The molecular phylogeny of *Alpinia* (Zingiberaceae): a complex and polyphyletic genus of gingers. Am. J. Bot. 92(1): 167-178.
- Kumar, G., Chauhan, B. and Ali, M. (2016). New alkadiene, benzyl linolenate and lawsone arabinosyl ester from the fruits of *Amomum subulatum* Roxb. J. Saudi Chem. Soc. 20: 476-479.
- Kusari, S., Hertweck, C. and Spiteller, M. (2012). Chemical ecology of endophytic fungi: origins of secondary metabolites. Chem. Biol. 19(7): 792-798.
- Leong-Skornickova, J., Sida, O., Jarolaimova, V., Sabu, M., Fer, T., Travnicek, P. and Suda, J. (2007). Chromosome numbers and genome size variation in Indian species of *Curcuma* (Zingiberaceae). Ann. Bot. 100(3): 505-526.
- List, P., 2010. The Plant List (www.theplantlist.org)
- Lutfia, A., Munir, E., Yurnaliza, Y. and Basyuni, M. (2021). Chemical analysis and anticancer activity of sesterterpenoid from an endophytic fungus *Hypomontagnella monticulosa* Zg15SU and its host *Zingiber griffithii* Baker. Heliyon 7(2): 06292.

- Maknoi, C. (2006). *Ph.D. Thesis.* Prince of Songkla University; Songkhla, Thailand. Taxonomy and Phylogeny of the Genus *Curcuma* L. (Zingiberaceae) with Particular Reference to Its Occurrence in Thailand.
- Munir, E. and Lutfia, A. (2019). Records of culturable endophytic fungi inhabiting rhizome of *Elettaria* in Hutansibayak, North Sumatera. *In*: IOP Conference Series: Earth and Environmental Science (Vol. 305, No. 1, p. 012004). IOP Publishing.
- Munir, E., Yurnaliza, Y., Lutfia, A. and Hartanto, A. (2021), June. Antifungal activity and IAA production by endophytic fungi isolated from *Elettaria* sp. *In*: IOP Conference Series: Earth and Environmental Science (Vol. 782, No. 4, p. 042037). IOP Publishing.
- My, V. T. N. and Thanh, N. V. (2020). Determination of structure and anticancer activity of MM 1 compound isolated from endophytic fungus Aspergillus terreus-RTN3 of Alpinia chinensis Rosc. In: International Conference on the Development of Biomedical Engineering in Vietnam (pp. 611-624). Springer, Cham.
- Oonmetta-aree, J. (2005). Effects of the Zingiberaceae spice extracts on growth and morphological changes of foodborne pathogens, Ph. D. thesis, School of Food Technology, Institute of Agricultural Technology, Suranaree University of Technology.
- Pansanit, A. and Pripdeevech, P. (2018). Antibacterial secondary metabolites from an endophytic fungus, *Arthrinium* sp. MFLUCC16-1053 isolated from *Zingiber cassumunar*. Mycology 9(4): 264-272.
- Praptiwi, K. D. P., Fathoni, A., Wulansari, D., Ilyas, M., Agusta, A. (2016). Evaluation of antibacterial and antioxidant activity of extracts of endophytic fungi isolated from Indonesian Zingiberaceous plants. NusantBiosci 8: 306–311.
- Ravindran, P. N. and Babu, K. N. eds. (2016). Ginger: the genus *Zingiber*. CRC press.
- Ravindran, P. N., Babu, K. N., Shiva, K. N. (2005). Botany and crop improvement of Ginger. *In*: Ravindran P N, Babu K N, editors. Ginger the genus *Zingiber*. Florida: CRC Press.15–85.
- Ravindran, P. N., Madhusoodanan, K. J. (2002). Cardamom: the genus *Elettaria*. London (UK): Taylor and Francis. pp. 368.
- Sabulal, B., Dan, M., Kurup, R., Pradeep, N. S., Valsamma, R. K. and George, V. (2006). Caryophyllene-rich rhizome oil of *Zingiber nimmonii* from South India: Chemical characterization and antimicrobial activity. Phytochemistry 67(22): 2469-2473.

- Schneider, P., Misiek, M. and Hoffmeister, D. (2008). *In vivo* and *in vitro* production options for fungal natural products. Mol. Pharm. 5: 234-242.
- Schulz, B., Boyle, C., Draeger, S., Römmert, A. K. and Krohn, K. (2002). Endophytic fungi: a source of novel biologically active secondary metabolites. Mycol. Res. 106(9): 996-1004.
- Septiana, E., Sukarno, N. and Simanjuntak, P. (2017). Endophytic fungi associated with turmeric (*Curcuma longa* L.) can inhibit histamine-forming bacteria in fish. Hayati J. Biosci. 24(1): 46-52.
- Shukla, Y., Singh, M. (2007). Cancer preventive properties of ginger: A brief review. Food Chem. Toxicol 45:683–690.
- Sieber, T. N. (2007). Endophytic fungi in forest trees: are they mutualists? Fungal Biol. Rev 21 (2–3): 75–89.
- Singh, D., Rathod, V., Ninganagouda, S., Herimath, J. and Kulkarni, P. (2013). Biosynthesis of silver nanoparticle by endophytic fungi *Pencillium* sp. isolated from *Curcuma longa* (turmeric) and its antibacterial activity against pathogenic gram-negative bacteria. J. Pharm. Res. 7(5): 448-453.
- Sivasothy, Y., Chong, W. K., Hamid, A., Eldeen, I. M., Sulaiman, S. F. and Awang, K. (2011). Essential oils of *Zingiber officinale* var. *rubrum* Theilade and their antibacterial activities. Food Chem. 124(2): 514-517.
- Smith, R. M. (1990). Alpinia (Zingiberaceae): a proposed new infrageneric classification. Edinb. J. Bot. 47(1): 1-75.
- Strobel, G. A. (2002). Rainforest endophytes and bioactive products. Crit. Rev. Biotechnol. 22(4): 315-333.
- Strobel, G. A. (2003). Endophytes as sources of bioactive products. Microbes Infect. 5(6): 535-544.
- Sunitha, V. H., Ramesha, A., Savitha, J. and Srinivas, C. (2012). Amylase production by endophytic fungi *Cylindrocephalum* sp. isolated from medicinal plant *Alpinia calcarata* (Haw.) Roscoe. Braz. J. Microbiol. 43: 1213-1221.
- Tan, R. X. and Zou, W. X. (2001). Endophytes: a rich source of functional metabolites. Nat. Prod. Rep. 18(4): 448-459.
- Ting, A. S., Meon, S., Kadir, J., Radu, S. and Singh, G. (2008). Endophytic microorganisms as potential growth promoters of banana. BioControl 53(3): 541-553.
- Uzma, F., Mohan, C. D., Hashem, A., Konappa, N. M., Rangappa, S., Kamath, P. V., Singh, B. P., Mudili, V., Gupta, V. K., Siddaiah, C. N. and Chowdappa, S. (2018). Endophytic fungi—alternative sources of cytotoxic compounds: a review. Front. Pharma col.9: 309.

- Venkateswarulu, N., Shameer, S., Bramhachari, P. V., Basha, S. T., Nagaraju, C. and Vijaya, T. (2018). Isolation and characterization of plumbagin (5-hydroxyl-2methylnaptalene-1, 4-dione) producing endophytic fungi *Cladosporium delicatulum* from endemic medicinal plants. Biotechnol. Rep. 20: 282.
- Vo, T. N. M., Nguyen, D. N. and Nguyen, V. T. (2020). Determination of structure and anticancer activity of MM2 compound, isolated from endophytic fungus *Aspergillus terreus*-RTN3 of *Alpinia chinensis* Rosc., IOP Conf. Series: Materials Science and Engineering 991: 012045. DOI:10.1088/1757-899X/991/1/012045.
- Wang, J. F., Liang, R., Liao, S. R., Yang, B., Tu, Z. C., Lin, X. P., Wang, B. G. and Liu, Y. (2017). Vaccinols J–S, ten new salicyloid derivatives from the marine mangrove-derived endophytic fungus *Pestalotiopsis vaccinni*. Fitoterapia 120: 164-170.
- Wiyakrutta, S., Sriubolmas, N., Panphut, W., Thongon, N., Danwisetkanjana, K., Ruangrungsi, N. and

Meevootisom, V. (2004). Endophytic fungi with antimicrobial, anti-cancer and anti-malarial activities isolated from Thai medicinal plants. World J. Microbiol. Biotechnol. 20(3): 265-272.

- Yang, X. and Eilerman, R.G. (1999). Pungent principal of *Alpinia galanga* (L.) Swartz and its applications. J. Agric. Food Chem. 47(4): 1657-1662.
- Yates, I. E., Bacon, C. W. and Hinton, D. M. (1997). Effects of endophytic infection by *Fusarium moniliforme* on corn growth and cellular morphology. Plant Dis. 81(7): 723-728.
- Yuan, Z. L., Zhang, C. L. and Lin, F. C. (2010). Role of diverse non-systemic fungal endophytes in plant performance and response to stress: Progress and approaches. J. Plant Growth Regul. 29(1): 116-126.
- Zhang, T. T., Lu, C. L. and Jiang, J. G. (2015). Antioxidant and anti-tumour evaluation of compounds identified from fruit of *Amonum tsaoko* Crevost *et* Lem. J. Funct. Food. 18: 423-431.